

Effect of Oxidation on the Quality of Biodiesel Produced from Nigerian Grown *Jatropha Curcas*

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Abstract— A lot of research work has proven that Nigerian *Jatropha curcas* is a potential feedstock for biodiesel production. However, the major drawback is its susceptibility to oxidation due to the presence of unsaturated fatty acids in the parent oil which adversely affects the quality of the biodiesel produced. This work investigated the effect of the oxidation on the quality of the biodiesel produced. Two important oxidation parameters: the acid value and the induction period were measured and correlated with biodiesel quality indices such as viscosity and cetane number. The results show that the Nigerian *Jatropha curcas* biodiesel (NJCB) fails to meet the minimum oxidation stability limit (IP of 6 hours) without antioxidant. Blending with petro-diesel as well as the addition of antioxidants leads to a composition having efficient and improved oxidation stability. The viscosity of all the samples are found to increase during the 24 weeks period of storage indicating an increase in the formation of oxidized products which lead to the formation of sediments and gum. The slight increase in the cetane number of the pure biodiesel and its blends can be attributed to the gradual saturation of the unsaturated methyl esters which are being oxidized into aldehydes and carboxylic acids.

Index Terms— biodiesel, oxidation, quality, blends, antioxidants.

I. INTRODUCTION

Biodiesel refers to a vegetable oil or animal fat-based diesel fuel consisting of long-chain alkyl esters or alkyl esters of fatty acids. It is a non-toxic, biodegradable and renewable fuel which can be used in compression ignition engines with little or no chemical modifications having significantly lower emissions compared to petro-diesel when it is burnt.

Furthermore, biodiesel does not contribute to the increase in carbon dioxide levels in the atmosphere and thus minimizes the intensity of the green house effect. It is also better than petro-diesel in terms of its properties such as sulphur content, flash point, aromatic content and biodegradability [12].

Biodiesel is produced through the transesterification of a range of vegetable feedstock oils or animal fats with alcohol; usually methanol to yield fatty acid methyl esters (FAME). Hence, the chemical composition of biodiesel can vary significantly resulting in extremely varied physical properties [13].

In spite of these advantages, stability is one of the major drawbacks for commercialization of such biodiesel [1]. Stability of biodiesel refers to the general resistance to change its fuel properties. It is desired that biodiesel resists the deteriorating effect of oxygen during storage over an extended period of time or due to elevated temperature. Oxidation stability, storage stability and thermal stability are used to describe oxidation affinity of biodiesel fuel in presence of oxygen [10]. It has been conclusively proved that susceptibility to oxidation or autooxidation during long term storage depends upon the composition of biodiesel [7].

Unsaturated fatty acids are significantly more prone to oxidation than saturated compounds. Also, polyunsaturated fatty esters are approximately twice as reactive to oxidation as monounsaturated esters [4]. Amongst the unsaturated fatty acids for example, linolenic acid exhibits the highest instability followed by linoleic and oleic acids. The Nigerian grown *jatropha curcas* seed oil shows a higher percentage of unsaturated fatty acids, largely linoleic and oleic (78.5%) which makes it liable to deterioration during storage over a long period.

This is attributed to the fact that these unsaturated fatty acid chains contain the most reactive sites, which are particularly susceptible to free-radical attack. The growth of oxidation process starts by reacting with oxygen through auto-oxidation mechanism, with the radical chain reaction following the steps of initiation, propagation, and termination [6]. During these reactions, several products such as peroxides, hydroperoxides, aldehydes, ketones, alcohols and some polymerization by-product are formed. Further, it is reported that the rate of oxidation of biodiesel was mainly influenced by environmental factors like ambient temperature, storage conditions and exposure of light [14].

The prevention of oxidation of biodiesel is difficult during natural storage condition, due to the fact that exposure to oxygen, light, heat and storage container are unavoidable during storage, handling and usage of biodiesel fuel [5]. However, some antioxidant additives could play a role to slow down the rate of oxidation. It is also stated that antioxidants inhibit the oxidation rate by slowing down the formation of free radicals. Careful selection of antioxidants is emphasized, because the improper type or concentration of antioxidant could act as pro-oxidant under certain condition [8].

The effects of various antioxidants on biodiesel oxidation stability have been investigated extensively by many researchers. Dunn (2008) showed that synthetic antioxidants such as tert-butylhydroxyquinone (TBHQ), butylated hydroxyanisole (BHA), tert-butylhydroxytoluene (BHT), pyrogallol (PY) and pyrogallate (PG) are generally more effective than natural ones (tocopherols). Many commercial additive formulations contain two or more antioxidants.

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Effective concentrations appear to be between 200 - 1000 ppm, depending on the feedstock and the type of stability test used. Chakraborty and Baruah (2012) found out that 1000 ppm of PG antioxidant was adequate to stabilize *Terminalia Belerica* biodiesel during 12 week storage period. Schober and Mittelbach (2004) showed that for a good antioxidant such as TBHQ, a 1000 ppm dose can improve biodiesel RIP by a factor of around 2, or even far more, depending on the substrate. TBHQ was again shown to be one of the most effective synthetic types, an initial dose of 400 ppm in undistilled RME achieved RIP 432 h after enduring 12 months of storage, compared to undistilled RME without any antioxidant which achieved just under 7h. Dunn (2008) presented a thorough review of such work, not covered in detail here. In summary, synthetic types, such as TBHQ, BHA, BHT and PG are generally more effective than natural types (tocopherols) so are generally preferred commercially.

II. METHODOLOGY

Collection and Preparation of *Jatropha Curcas* Seed Oil

Jatropha curcas seed oil was obtained from National Research Institute for Chemical Technology (NARICT), Zaria, Nigeria. The crude *Jatropha curcas* seed oil was analyzed in terms of its physiochemical properties in accordance with American Society of Testing Material (ASTM) standard.

Analysis of *Jatropha Curcas* Seed Oil

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Determination of Acid Value

The acid value is defined as the mg of potassium hydroxide necessary to neutralize fatty acids in 1g of sample and is measured as per AOCS Method Ia - 64. It reflects the amount of free fatty acid content in *Jatropha* oil. The indicator solution (1.0% phenolphthalein indicator in isopropyl alcohol) was added to the required amount of solvent (equal parts by volume of isopropyl alcohol and toluene) in ratio of 2ml to 125ml and neutralized with alkali to a faint but permanent pink colour. 5g of the well mixed sample was weighed into an Erlenmeyer flask and 125ml of the neutralized solvent mixture was added. It was well shaken and ensured that the sample was completely dissolved before titration. The sample was shaken vigorously while titrating with the standard alkali (0.1M KOH) until the first permanent pink colour was observed. The acid value was calculated as follows:

$$\text{Acid value} = \frac{\text{Titre value} \times \text{Normality of KOH}}{\text{Mass of Jatropha oil}} \quad (1)$$

Determination of Viscosity

The viscosity of *Jatropha* oil was measured at 24°C using a digital rotary viscometer. The sample cup was removed and the spindle CP-40 was carefully attached to the lower shaft and inserted into the cup containing 25g of the oil sample. The motor switch was turned "on" and rotated at a speed of 60 rpm and sufficient time was allowed for the display reading to stabilize.

Determination of Free Fatty Acid

The method employed for this analysis was AOCS method Ca-5a-40. It gives the percentage by weight of specified fatty acid present in the oil. 5g of the *Jatropha* oil was weighed into 250 ml conical flask. 25 ml diethyl ether with 25 ml ethanol mixture was added to the *Jatropha* oil. The mixture was boiled on a hot plate until all the oil dissolved completely. 3 drops of indicator phenolphthalein was added to the mixture and titrated with 0.1 M sodium hydroxide with constant shaking until a pink colour persisted for 30 minutes. The percentage of FFA was calculated using Equation 2.

$$\% \text{ FFA} = \frac{\text{Titre value} \times \text{Normality of KOH} \times 56.1}{\text{Mass of Jatropha oil}} \quad (2)$$

Esterification of the *jatropha curcas* oil

The oil was esterified to reduce the percentage of free fatty acid in it. The esterification process was carried out using methanol and sulphuric acid catalyst. About 1000 g of the *Jatropha* oil was heated to a temperature of 60 °C in a conical flask and a mixture of methanol (30% by weight of oil) and sulphuric acid (1% by weight of oil) was added to it. The resulting mixture was then heated but maintained at a temperature of 60 °C with stirring at 600 rpm for one hour.

Transesterification of the *jatropha curcas* oil to Biodiesel

The standard alkali-catalyzed transesterification process was used for the conversion of the *Jatropha* oil to biodiesel. The transesterification process was carried out using methanol and potassium as catalyst.

500 g of the esterified oil was heated to a temperature of 60 °C in a conical flask and a mixture of methanol and calcium oxide catalyst (8% by weight of oil) was added to it. The resulting mixture was then heated but maintained at a temperature of 60 °C with stirring at 700 rpm for one hour.

The mixture was thereafter poured into a separating funnel where the biodiesel and glycerol were separated on standing. The solid catalyst was recovered by filtration.

Blend Preparation

The *Jatropha curcas* biodiesel and petro-diesel were mixed in specified proportions (v/v) to obtain the blends. For example, 10% biodiesel was mixed with 90% petro-diesel to obtain B10. Similarly, B20 and B30 were also prepared.

Evaluation of Fuel Properties

The major fuel quality parameters viscosity, acid value, peroxide value and induction period of the pure *Jatropha curcas* biodiesel and its blends with petro-diesel with and without antioxidants, were determined during 24 weeks of storage at periodic intervals.

III. RESULTS

a. Analysis of Crude *Jatropha curcas* Oil

The results of crude *Jatropha* oil analysis are presented in Table 1 These show that the properties of *Jatropha* seed oil used in this work are in conformity with literature values. The oil sample has a high value of FFA (16.4%) much higher than the 1% specified limit, hence the oil needs to be esterified before transesterification. High value of FFA > 1% adversely affect the transesterification reaction in which the FFA in the oil reacts with the alkali catalyst to produce soap (saponification) hence decreasing the catalyst amount needed for the transesterification reaction [11].

Table 1: Properties of the crude *Jatropha curcas* seed oil

Parameter	Unit	Value	Test method	ASTMD Standard
Density	kg/m ³	872	ASTMD 287	860 – 933
Viscosity @ 40 °C	mm ² /s	4.3	ASTM D6751	4.0 – 6.0
AV			AOCS Te 1a-64	
FFA	mgKOH/kg	0.97	AOCS(Ca5a-40)	0.92 – 6.16
Moisture content	%	16.4	Karl Fisher	-
		7.5		

Analysis of the fatty acid composition (Table 2) of the crude *Jatropha* oil indicates that the oil consisted principally of linoleic acid (40.39%) followed by oleic acid (38.14%), palmitic acid (14.43%) and stearic acid (7.04 %). This is in conformity with that reported by Siddharth and Sharma (2011). It indicates that the oil contains more unsaturated fatty acids (linoleic acid and oleic acid) than the saturated fatty acids (palmitic acid and stearic acid). The oxidative stability of biodiesel is a function of the fatty acid composition of the parent oil and decreases with larger amount of unsaturated fatty acids in the parent oil.

Table 2: Free fatty acid composition of the *Jatropha* oil

Fatty Acid	Value (wt. %)
Palmitic acid (16:0)	14.43

Stearic acid (18:0)	7.04
Oleic acid (18:1)	38.14
Linoleic acid (18:2)	40.39
Linolenic (18:3)	-
Saturates	21.47
Unsaturates	78.53

Evaluation of Fuel Properties of Fresh *Jatropha Curcas* Biodiesel

The fuel properties of freshly prepared *Jatropha* biodiesel and its blends in comparison to petro-diesel are presented in Table 3. It is found that the fuel properties of *jatropha curcas* biodiesel and its blends satisfy biodiesel standards. (EN 4214 and ASTM D6751)

Table 3: Fuel properties of *Jatropha* biodiesel and blends

Parameter	Unit	B100	B10	B20	B30	ASTMD Standard	Petro-diesel
Density	kg/m ³	872	826	832	843	860 – 933	835.12
Viscosity	mm ² /s	4.35	2.77	3.34	3.52	1.9 – 6.0	2.78
AV	mgKOH/g	0.35	0.22	0.27	0.28	0.5 (max)	0.10
PV	mg/kg	4.71	2.99	3.61	3.81		-
IP	Hours	3.31	26	22	17	6(B100) 20(blend)	-

Changes in Fuel Properties of *Jatropha Curcas* Biodiesel and its Blends on Storage

The change in fuel quality during storage due to oxidation is evaluated through monitoring the changes in viscosity (a physical property), acid value (based on secondary oxidation

products), peroxide value (based on primary oxidation products), Rancimat induction period (based on accelerated oxidation) and cetane number.

The quality deterioration regarding these specific parameters up to 24 weeks of storage is presented below.

Viscosity

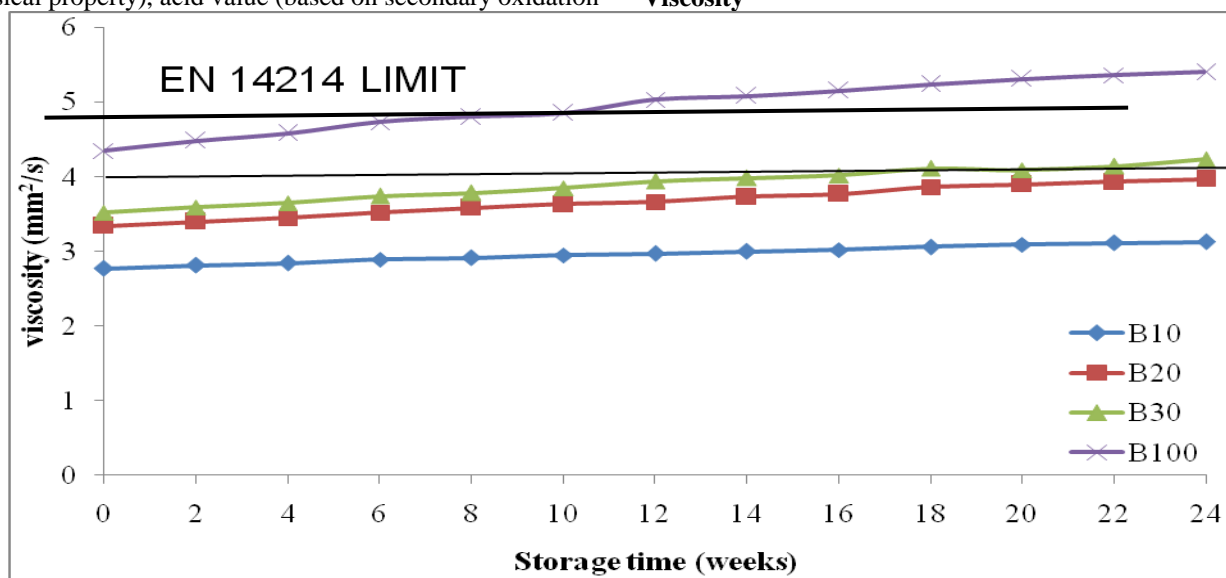


Fig. 3.1 Variation in the viscosity of blend samples with storage period

The variations in viscosity with storage period are presented in Fig. 4.1 for all the samples under investigation. Generally,

the viscosity values of all the samples are found to increase with period of storage. This can be attributed to the formation

of oxidized products which lead to the formation of sediments and gum [3].

The rates of increase in viscosity of the samples however are not uniform. Pure biodiesel (B100) which increased from 4.35 mm²/s to 5.40 mm²/s during the 24 weeks shows a higher rate of increase compared to the blended samples.

Chemical reaction and the formation of products during interaction of biodiesel and petro-diesel is a likely reason for

the different rates of increase in viscosity amongst the samples.

Viscosity of B100 is above that of petro-diesel throughout the 24 week storage period while that of B30 goes beyond the viscosity of petro-diesel after the 16th week of storage.

All the samples are within the EN 14214 limit of 5 mm²/s up to 11 weeks but B100 goes above the minimum limit after the 11th week.

Acid value

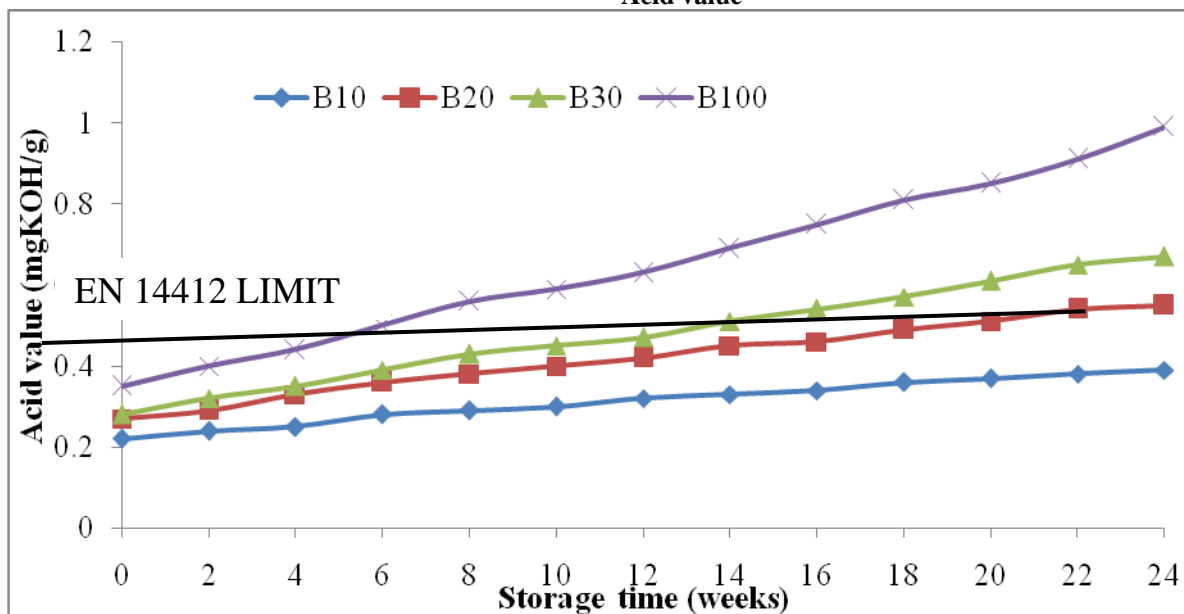


Fig. 3.2 Variation in acid value of blend samples with storage period

The variations in acid value (AV) with storage period are presented in Fig. 3.2 for all the samples under investigation. The results show an increase in AV for all the samples.

Similar rate of increase in AV is observed for the two blend samples B30 and B20. The rate of increase in the AV of B10 during storage is the least of all the samples while pure biodiesel B100 exhibited the highest rate of increase from 0.35 mgKOH/g to 0.99 mgKOH/g during the 24-week storage period. The AV for B100 is within the EN 14412 limit of 0.5 mgKOH/g (max) up to 7 weeks; B30 up to 16 weeks; B20 up to 24 weeks; B10 within limit throughout the 24 week storage period.

During storage, the esters are first oxidized to form peroxides, which then undergo complex reaction and further oxidize into acids, and hence increase the acid value. Also, the hydrolysis of esters into alcohol and acid by the traces of water present in the sample increases the acid value (Mittelbach, 2004)

Effect of Antioxidant Additives on Storage of *Jatropha Curcas* Biodiesel and its Blends

Further investigation was carried out to check the effectiveness of pyrogallol (PY) antioxidant additives on the pure biodiesel and its blends. PY antioxidant is found to be the most effective antioxidant for *jatropha curcas* biodiesel

[[15]. The different samples were doped with varying concentrations of PY antioxidant of 50 ppm, 100 ppm and 150 ppm.

Effect of PY Antioxidant on the Viscosity of Samples

Fig. 3.3 shows the effect of PY antioxidant on the viscosity of *jatropha curcas* biodiesel and its blends at varying concentrations. The viscosity of pure *jatropha* biodiesel without antioxidants is found to increase by 24% while the viscosity of the blends B20 and B30 showed a lower rate of increase of 20% and 19% respectively over a period of 24 weeks. The addition of PY antioxidant however slowed down the rate of increase in viscosity in all the three samples due to deceleration in the oxidation process, which in turn decreases the peroxide formation.

Without antioxidants, the viscosity of the pure biodiesel B100 goes above the 5mm²/s EN 14214 limit while the viscosity of the blends B10, B20 and B30 fall within the limit. Application of antioxidants suppresses the viscosity of all the samples and this increases with the quantity of antioxidant. Antioxidant dose of 100 ppm and 150 ppm exhibited similar rate of retarding the rate of increase in viscosity for all the samples.

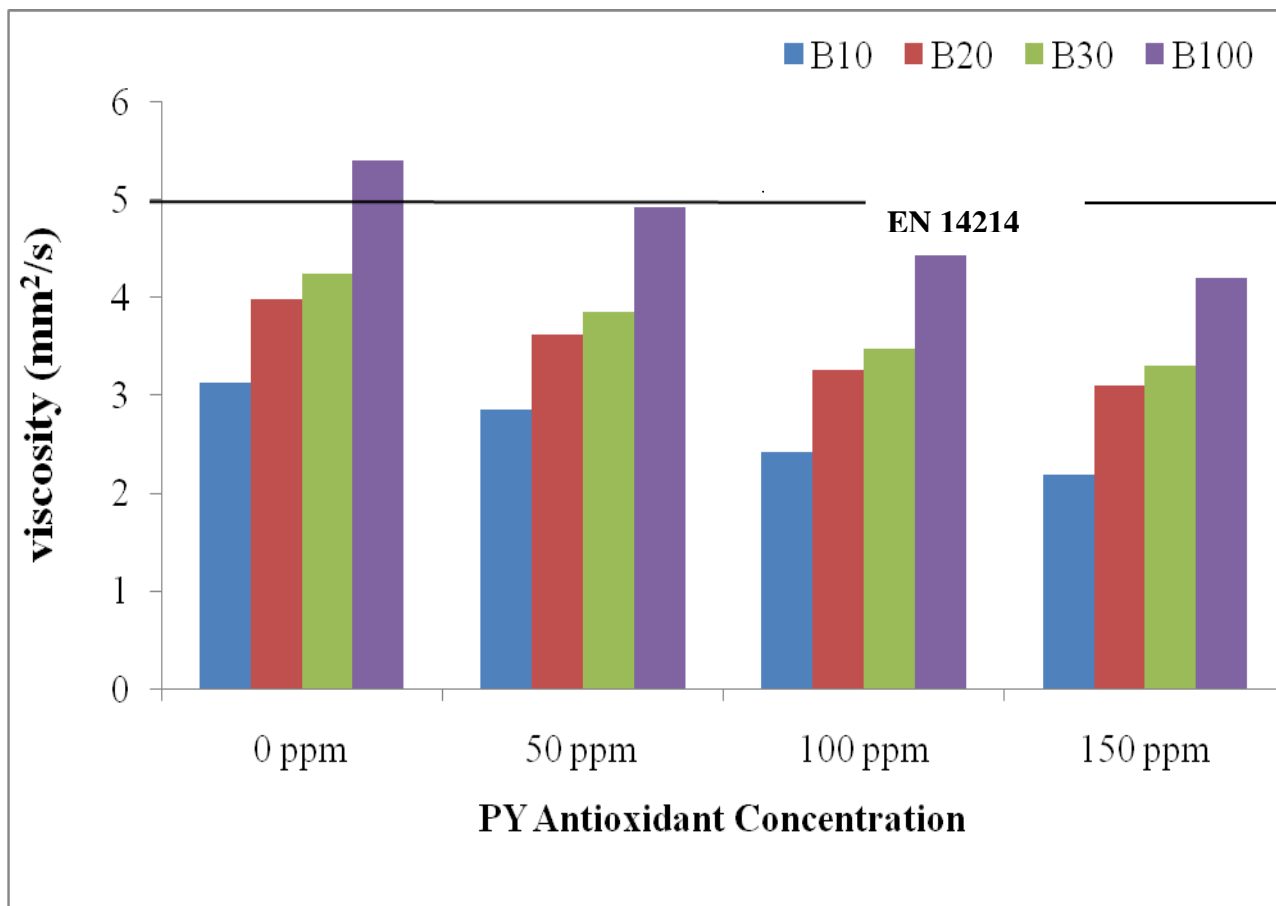


Fig. 3.3 Effect of PY antioxidant on the viscosity of pure *Jatropha* biodiesel and blends

Effect of PY Antioxidant on the acid value of Samples

Fig. 3.3 shows the effect of PY antioxidant on the acid value of *Jatropha curcas* biodiesel and its blends at different concentrations. The AV of pure *Jatropha* biodiesel without antioxidant increased from 0.35 mgKOH/g to 0.99 mgKOH/g over a storage period of 24 weeks. The acid values of the blends B20 and B30 without antioxidant exceeded the maximum limit of 0.5 mgKOH/g after the 20th and 16th weeks

respectively. The addition of PY antioxidant significantly decreased the rate of increased in acid value of all the samples due to deceleration in the oxidation process.

Without antioxidant, only B10 falls below the minimum 0.5 mgKOH/g set by EN 14412. Application of PY antioxidant decreases the acid value of all samples, indicating a reduction in oxidation. 50 ppm is adequate for B20, blend B30 requires up to 150 ppm while B10 does not require any antioxidant. B100 requires more than 150 ppm.

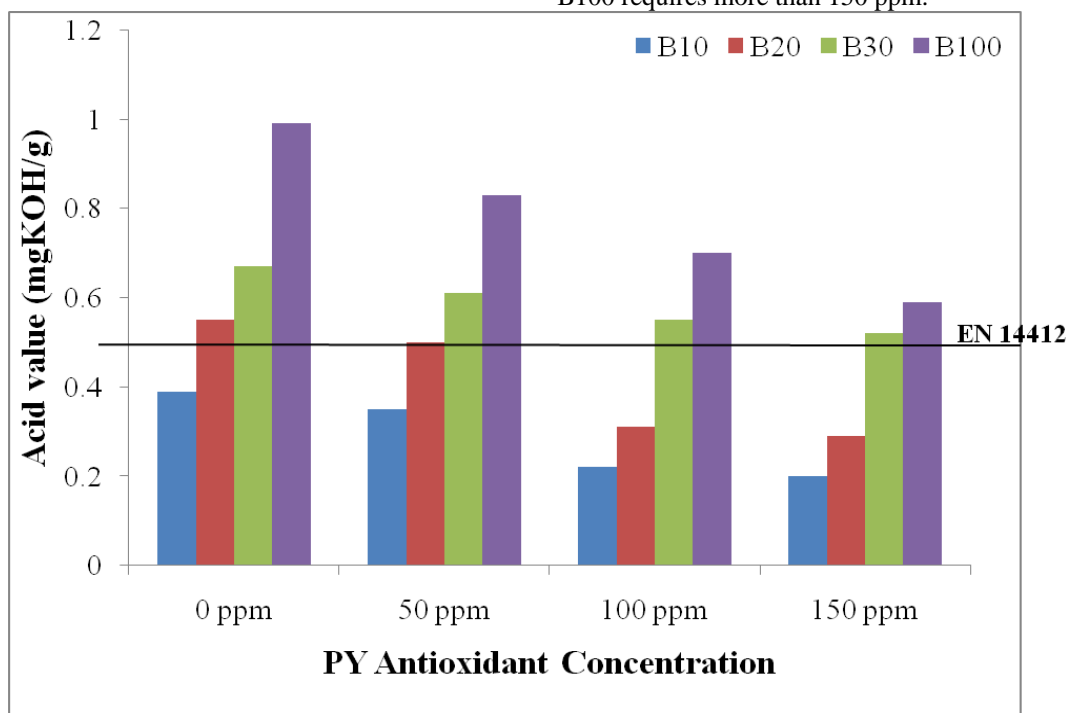


Fig. 3.4 Effect of PY antioxidant on the acid value of *Jatropha* biodiesel and blends

Effect of PY Antioxidant on the induction period of Samples
 Fig. 3.5a and 3.5b show the effect of PY anti-oxidant concentration on the induction period of pure jatropha biodiesel and its blends.
 Without antioxidant, the induction period (IP) of 3.31 hours for the pure biodiesel B100, falls short of the minimum IP of 6 hours set by EN 1214.

Without antioxidant, both B10 (IP 26.17 hours) and B20 (IP 22.23 hours) go above the minimum of 20 hours set by EN 14214 while B30 (IP 17.13 hours) falls below the 20 hours standard. The IP increases with increase in antioxidant concentration.

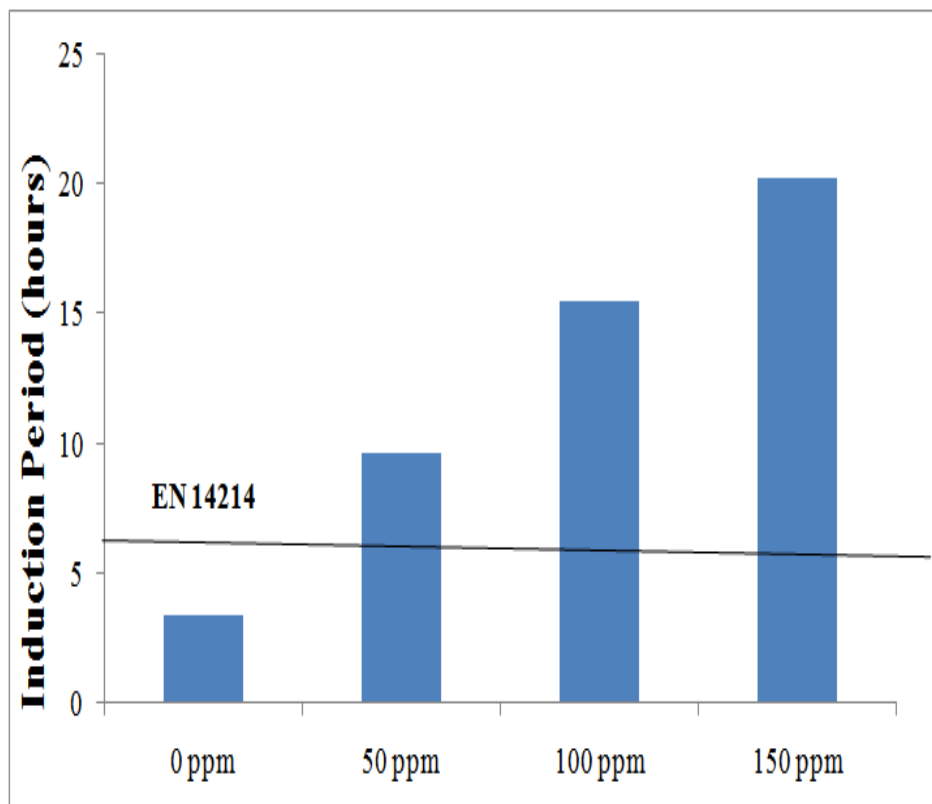


Fig. 3.5a Effect of PY antioxidant on the induction period of pure *Jatropha* biodiesel

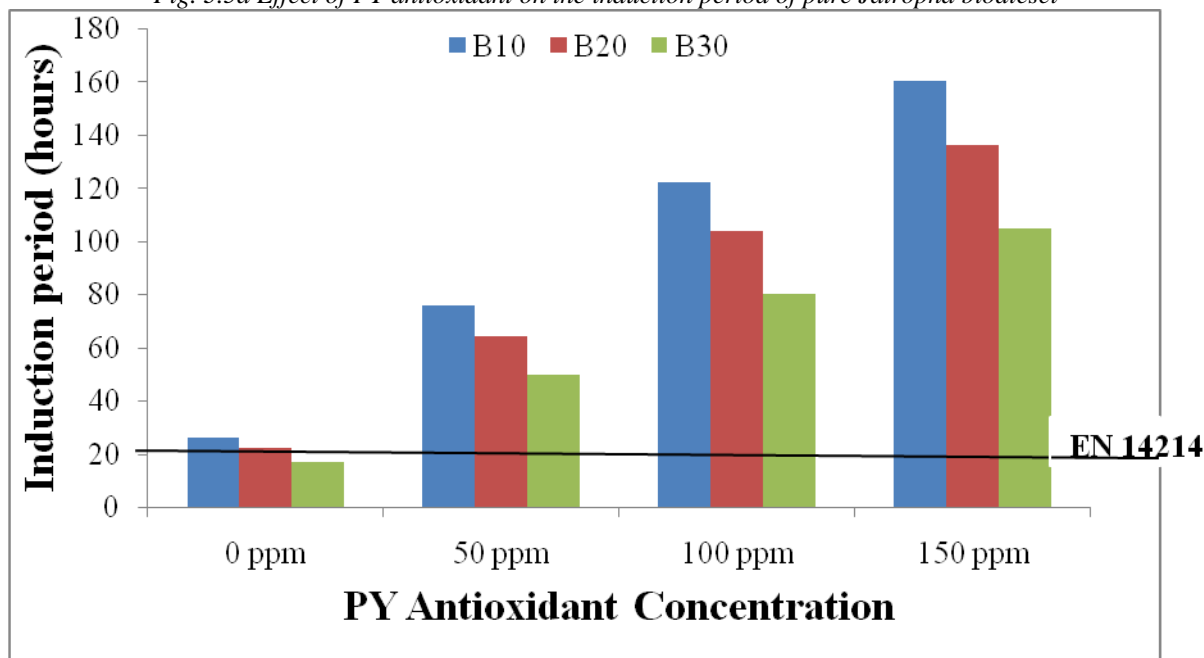


Fig. 3.5b Effect of PY antioxidant on the induction period of *Jatropha* biodiesel blends
 Cetane Number

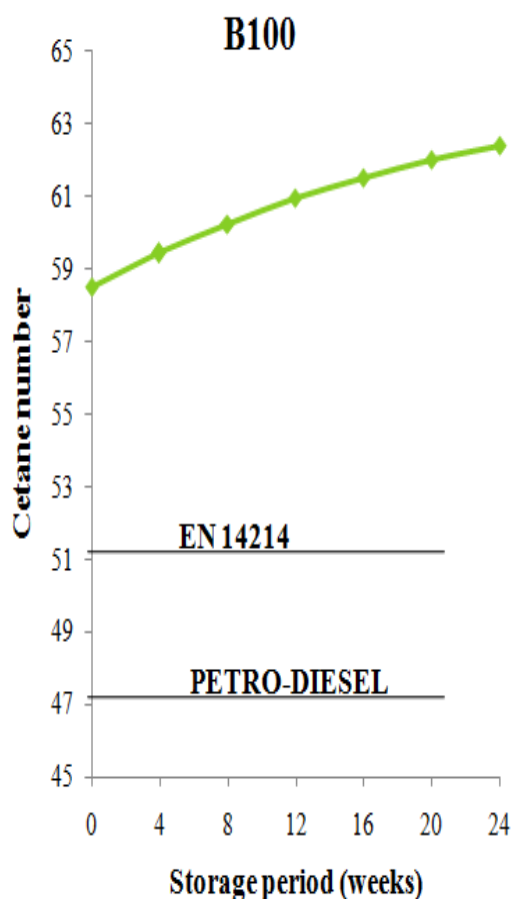


Fig. 3.6a Variation of cetane number of B100 with storage period

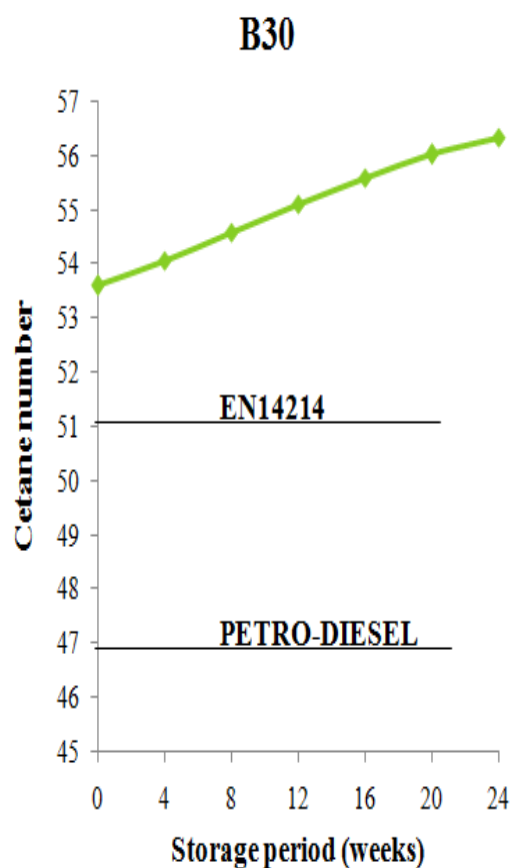


Fig. 3.6b Variation of cetane number of B30 with storage period

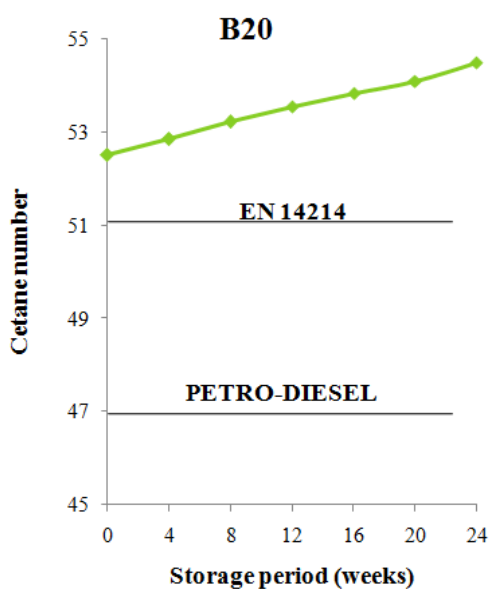


Fig. 3.6c Variation of cetane number of B20 with storage period

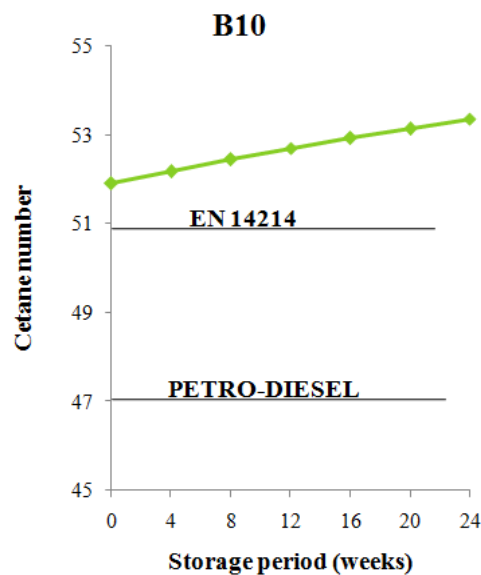


Fig. 3.6d Variation of cetane number of B10 with storage period

Fig. 3.6 a – d show the variation of cetane number (CN) of pure biodiesel B100 as well as the biodiesel blends B10, B20 and B30 during the 24 weeks period of storage. The results show a small and gradual increase in cetane number for all the samples and are above that of petro-diesel as well as that of the CN ≥ 51 set by EN 14214.

CN for B100 increased from 58.5 to 62.4; that of B30 increased from 53.6 to 56.3; CN for B20 increased from 52.5 to 54.5 while CN for B10 increased from 51.9 to 53.4. Knothe (2005) established that cetane number decreases with increasing unsaturation and increases with increasing chain length. Hence the slight increase in the cetane number of the

pure biodiesel and its blends can be attributed to the gradual saturation of the unsaturated methyl esters which are being oxidized into aldehydes and carboxylic acids.

IV. CONCLUSION

The conclusions derived from this study are summarized as follows:

The quality of biodiesel produced from Nigerian grown *Jatropha curcas* seed oil is found to be affected adversely by oxidation. Even though the cetane number shows slight increase during the storage period, the increase in the viscosity and acid value is an indication of accumulation of oxidation products such as gums and sediments which negatively affect the engine. Blending and the addition of PY antioxidant were effective in slowing down oxidation and thus reduce the negative effect on the quality of the biodiesel.

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